

Research Article

Analysis of Copy Number Variations and Knockdown in Zebrafish Pronephros Identifies Novel Candidate CAKUT Genes

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Abstract

Congenital anomalies of the kidney and urinary tract (CAKUT) are serious birth defects that occur in ~1:1000 pregnancies. Mutations in ~40 different genes are likely to account for these disorders. However, because mutations in unique genes affect a small number of patients with variable penetrance and expressivity, identification of causative genes has been challenging. We identified six novel candidate CAKUT genes in regions of genomic imbalance and showed pronephric phenotypes when gene expression was reduced in zebrafish.

Introduction

Congenital anomalies of the kidney and urinary tract (CAKUT) are the most common cause of chronic kidney disease in children. They account for ~48-59% of childhood chronic kidney disease (CKD) and 34-43% of childhood end stage kidney failure requiring dialysis and transplantation [1]. CKD in infants and children is associated with serious sequelae, including reduced life expectancy, cardiovascular disease, impaired growth and neurocognitive delay. Genetic variants contribute significantly to the pathogenesis of CAKUT [2]. Syndromic forms of CAKUT, often with extra-renal manifestations are typically monogenic disorders with high penetrance that are readily diagnosable. More challenging is identifying the genetic basis for the more common sporadic forms of CAKUT because of the high degree of locus and allelic heterogeneity, reduced penetrance and variable severity. To date, ~40 genes have been implicated in sporadic, non-syndromic CAKUT. However, this only accounts for ~25% of CAKUT cases, indicating that many more genes are expected to contribute to this developmental disorder [3, 4]. Moreover, in many of these reports there are no functional data to support the pathogenicity of the candidate gene variants. It has recently been appreciated that 10-17% of CAKUT cases are attributed to copy number variations (CNVs) [5, 6]. Genes contained in these regions of chromosomal imbalance represent novel genetic causes of CAKUT. Chromosomal microarray analyses were used to identify genomic imbalances (deletions or duplications) in two cohorts of children with CAKUT [7]. Here we report results of functional analysis of 12 novel candidate CAKUT genes within regions affected by these structural variants.

Results and Discussion

We analyzed a dataset that identified CNVs in a cohort of 457 CAKUT patients, but were extremely rare or absent in several control cohorts totaling 11,787 individuals. CNVs can identify dosage-sensitive genes that are linked to phenotypes. We used the following criteria to prioritize genes to test in functional assays: expression in the mouse urogenital tract in public databases (GUDMAP, Geo) or our own studies in the mouse embryonic kidney; functional data implicating the gene in kidney formation in a model organism; a biological pathway with a strong link to kidney development. We also considered whether mutations in the gene were associated with a human congenital anomaly syndrome, with or without known urogenital tract anomalies.

We tested whether knockdown of genes disrupted by these rare CNVs in the CAKUT cohort affected formation of the pronephros in zebrafish. We queried the Zebrafish Model Organism Database (ZFIN) to identify orthologs of candidate human CAKUT genes contained within regions of genomic imbalance. We used morpholinos to test if gene knockdown affected pronephric development in transgenic fish that expressed GFP in the glomerulus [Tg(wt1b:egfp)^{li2}] and the pronephric duct [Tg(cdh17:egfp)^{pi305}]. Single cell embryos were injected with morpholino oligonucleotide at 0.0125-0.25 mM and were visualized by epifluorescent microscopy at 48 hours post-fertilization.

Out of twelve genes tested, knockdown of six genes showed a pronephric phenotype (table 1). PCDH15 encodes for a member of the Protocadherin protein family. The gene is mutated in Usher

syndrome type 1D/F, which is associated with sensorineural hearing loss and retinitis pigmentosa (OMIM #601067). Studies of the Usher syndrome protein network has revealed important molecular links to ciliopathies, many of which are associated with nephronophthisis, a common cause of childhood chronic kidney disease [8]. HACE1 is a HECT-domain and ankyrin repeat-containing E3 ubiquitin ligase. Homozygous loss of function mutations lead to spastic paraplegia and neuro developmental delay (OMIM #616756). The gene is highly expressed in fetal kidney and its loss of expression may play a role in the pathogenesis of sporadic Wilms tumor [9, 10]. *Slc8a1a* encodes for a sodium calcium exchanger. Knockdown of the gene in renal epithelial cells destabilized E-cadherin and disrupted canonical Wnt signaling, thereby affecting the mesenchymal to epithelial transition, an essential step in formation of the kidney [11]. *Lrp1b* encodes for the low-density lipoprotein receptor related-protein 1b. Variants of this this gene are associated with insulin resistance and childhood BMI [12-14]. It has been suggested that maternal hyperglycemia in gestational diabetes results alters DNA methylation at this locus and thereby contributes to fetal metabolic reprogramming [15]. Therefore, *LRP1B* may be a candidate gene involved in gene-environment interactions in conditions such as diabetes, which are associated with

a higher risk of birth defects. In addition, deletion of *LRP1B* has been observed in adult Wilms tumor [16].

Two of the genes were studied in more detail because they are both inhibitors of receptor tyrosine kinase signaling, a pathway that is critical for kidney development in mice and humans [17]. Knockdown of *Spred1* and *Sprouty2* (*Spry2*) produced similar, dosage-sensitive phenotypes with two independent morpholinos. The observed phenotype included glomerular cysts and lack of extension of the pronephric duct leading to the absence of a patent opening at the cloaca (Figures 1, 2). Defective growth and branching of the nephric duct and ureteric bud are characteristic of mutations in the c-Ret receptor tyrosine kinase, which is essential for normal development of the mammalian kidney and lower urinary tract [17]. *Spry2* plays a critical role in regulating c-Ret in developing kidney [18, 19]. *Spred1* encoding for the *Sprouty1*-related gene product is a negative regulator of Ras-Mitogen Activated Protein Kinase (MAPK) activity. Point mutations in c-RET that disrupt MAPK signaling lead to congenital anomalies affecting the kidney and lower urinary tract [20]. Mutations in *SPRED1* causes Neurofibromatosis type I (Legius syndrome) which is associated with childhood renal cancer (OMIM#611431).

Table 1 :

Morpholino	Injection []	fish tg line	n injected	n affected	% phenotype	phenotype description
Spry2 #1	0.25mM	<i>Cdh17</i>	61	30	49.2	CD, TR
Spry2 #1	0.125mM	<i>Cdh17 and Wt1b</i>	180	26	14.4	CD, GC
Spry2 #2	0.25mM	<i>Cdh17 and Wt1b</i>	182	30	16.5	CD, GC, GM
Spred1 #1	0.125mM	<i>Cdh17 and Wt1b</i>	44	11	25.0	CD, GC, GM
Spred1 #2	0.125mM	<i>Cdh17 and Wt1b</i>	12	2	16.7	CD
Spred1 #2	0.0625mM	<i>Wt1b</i>	47	7	14.9	GC, SD
Spred1 #2	0.0125mM	<i>Cdh17 and Wt1b</i>	33	17	51.5	GC, GM
<i>Pcdh15</i>	0.25mM	<i>Cdh17 and Wt1b</i>	182	39	21.4	GC, TR, OT, SD
<i>Hace1</i>	0.25mM	<i>Cdh17 and Wt1b</i>	79	9	11.4	GC, GM
<i>Lrp1b</i>	0.25mM	<i>Cdh17 and Wt1b</i>	23	17	73.9	CD, GC, SD
<i>Lrp1b</i>	0.125mM	<i>Cdh17 and Wt1b</i>	3A	13	38.2	CD, SD
<i>Slc8a1a</i>	0.25mM	<i>Cdh17 and Wt1b</i>	44	2	4.5	GC

CD (tubules don't exit fish at cloacal duct), GC (glomerular cyst), GM(glomerular malformation), SD(severe developmental defect), TR(truncated tubule), OT(obstructed/enlarged tubule)



Figure 1. A. Low power images of pronephric phenotypes. Bi-transgenic fish expressing GFP under the control of the *Wt1* promoter in the glomerulus and in the pronephric duct under control of the *Cdh17* promoter. Uninjected fish displayed normal formation of the glomerulus and pronephric duct. B. Morpholino knockdown of *Spred1* (*Spred1* MO) resulted in glomerular cyst seen in *Wt1* transgenic mice. C. Morpholino knockdown of *Sprouty2* (*Spry2* MO) resulted in a shortened pronephric duct in *Cdh17* transgenic fish.

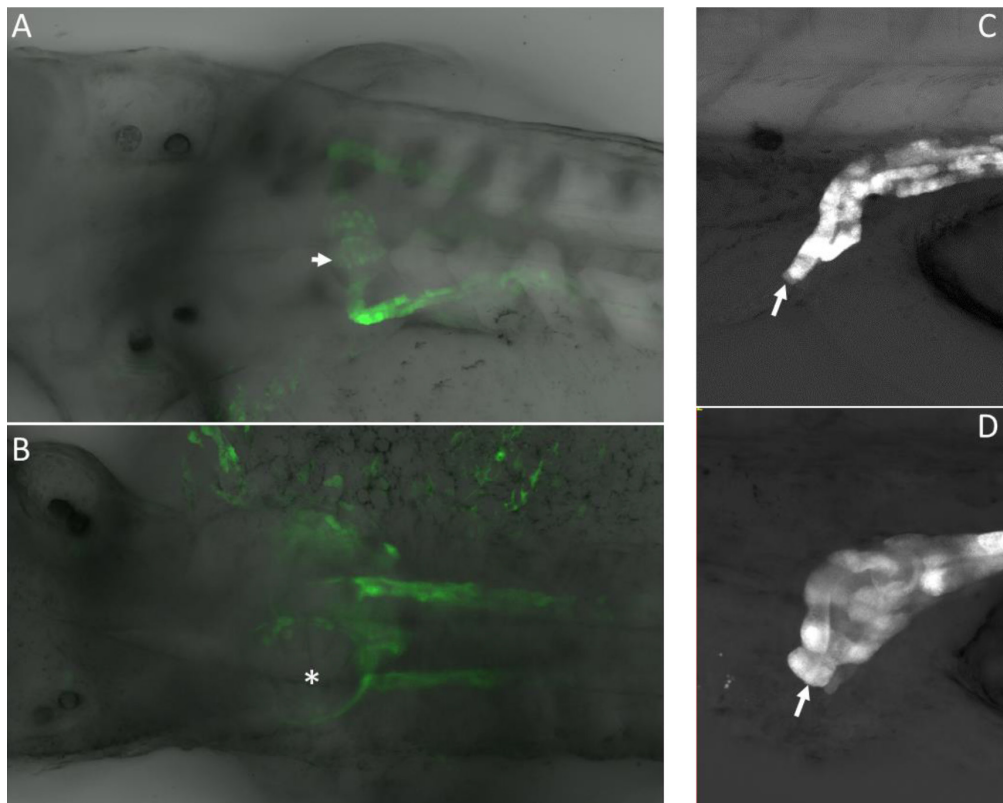


Figure 2. A. High power confocal images of pronephric phenotypes. Normal glomerulus in a control (uninjected) embryo that expressed GFP from the *Wt1* promoter. B. *Spred1* morpholino knockdown caused glomerular cyst formation (asterisk). C. Pronephric duct shown exiting at the cloaca (arrow). D. Blunted pronephric duct that fails to exit at the cloaca due to morpholino knockdown of *Spry2* (arrow). Note the dilatation at the distal end of the pronephric duct that occurred because the duct is not patent.

In conclusion, we have identified six novel candidate CAKUT genes by combining CNV data and functional analysis in zebrafish. Validation of these genes as causative of human CAKUT awaits discovery of additional affected individuals with mutations in these genes using whole exome sequencing.

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